Chemical Synthesis of 2-O-(α-L-Fucopyranosyl)-3-O-(2-acetamido-2-deoxy-α-Dgalactopyranosyl)-D-galactose, the Terminal Structure in the Blood-group A Antigenic Determinant

By Serge David,* Andre Lubineau, and Jean-Michel Vatèle

(Laboratoire de Chimie Organique Multifonctionnelle, Université de Paris-Sud, 91405 Orsay Cedex, France)

Summary The trisaccharide (1), the terminal structure in the blood-group A antigenic determinant, has been prepared from the trisaccharide (4) which was obtained from the product of a cycloaddition and seems to be a suitable precursor for synthesis of modified A and B blood-group antigens.

BLOOD-GROUP antigens from erythrocytes membranes are glycolipids which carry the A-, B-, H-, and Le-determinants

on the outer part of their oligosaccharide chains. The terminal structures of the A and B antigens, the trisaccharides (1) and (2), differ only at C-2 of one of the non-reducing units.¹ The MeCO group in the trisaccharide (1) seems most important in this connection since treatment of normal A_1 erythrocytes with a bacterial N-acetylgalactosamine deacetylase leads to a pH-dependent B specificity, probably due to structure (3).² It was thus of interest to test modified A(B) trisaccharides as inhibitors in these systems,

and we have prepared as precursor for such syntheses the trisaccharide (4), the usefulness of which is demonstrated by its conversion into the A determinant (1). We are not aware of any published synthesis of (1), but a preparation by coupling of a 2-azido-2-deoxy derivative³ has been disclosed at a meeting.⁴



Bn = benzyl; Men = (-)-menthyl; All = allyl; Tr = trityl.

Acetal formation (PhCHO, ZnCl₂) with benzyl 2-O-allyl- α -D-galactopyranoside⁵ gave the starting material, benzyl 2-O-allyl-4,6-O-benzylidene- α -D-galactopyranoside (5) (86%) [m.p. 116 °C (from ether-light petroleum); $\alpha_D^{20} + 141^\circ$].† From the alcohol (5), by repetition of the described reaction sequence⁶ were prepared in succession the trans enynyl ether (6) $(36\%)^+_+$ [eluent, chloroform-ether-light petroleum (1:4:5); m.p. 126 °C (from ether-light petroleum; $\alpha_{\rm p}^{20}$ $+211^{\circ}$), the trans dienvel ether (7) (90%) [eluent, ether-light petroleum (1:1); oil; $\alpha_D^{20} + 195^\circ$], and then after cycloaddition with (-)-menthyl glyoxylate, a mixture of dihydropyrans (85%). Isomerization (0.1 ml of BF₃-Et₂O in 20 ml of ether per g of mixture) gave the 'a-D' compound (8) (43%) [eluent, toluene-ether-light petroleum (80:20:15)oil; $\alpha_D^{20} + 57^\circ$] the configuration of which was ascertained by the described procedure.⁶ The allyl group was removed⁷ and the resulting disaccharide (9) (62%) [eluent, ether-light petroleum (1:1); m.p. 137 °C; $\alpha_D^{20} + 64^\circ$] was fucosylated on its free OH by the common-ion procedure.⁸ The protected trisaccharide (10) (85%) [eluent, toluene-etherlight petroleum (8:2:3); oil, $\alpha_D^{20} - 41^\circ$] was reduced with $LiAlH_4$ to the primary alcohol (4) (90%) [eluent, etherlight petroleum (4:1); m.p. 140 °C (from ether-light petroleum; $\alpha_{\mathbf{p}}^{20} + 15.6^{\circ}$].

The trisaccharide (4), which can be readily prepared in 10 g amounts, is a convenient precursor for the synthesis of modified A, B determinants. Extension of known procedures^{9,10} gave the *D-lyxo* epoxide (11) (74%) (eluent, ether; foam; $\alpha_{20}^{20} + 38^{\circ}$), the allylic alcohol (12) (74%) [eluent, ether-methanol (95:5); foam; $\alpha_{20}^{20} + 58^{\circ}$], the trityl ether (13) (94%) [eluent, ether-light petroleum (1:1); powder; $\alpha_{20}^{20} + 4^{\circ}$] and the unsaturated ketone (14) (85%) [eluent, chloroform-ether-hexane (4:1:4); m.p. 106 °C (from ether); $\alpha_{20}^{20} + 5\cdot5^{\circ}$].

Without isolation of intermediates, the ketone (14) was treated in succession with OsO4 (1.1 equiv. in pyridine), hydroxylamine hydrochloride, and then acetic anhydride, to give the NOO-triacetate (15) (68%) [eluent, chloroform ether (20:3); amorphous; $\alpha_{\rm D}^{20} + 18^{\circ}$]. LiAlH₄ reduction of (15), followed by selective N-acetylation in methanol, gave a mixture (58%) of talo (17%) and galacto (83%) compounds. The latter, a foam, was separated by chromatography (ethermethanol, 95:5), and the protective groups were removed by hydrogenolysis [1 week at room temperature in waterethanol (5:95) solution, in the presence of an equal weight of 10% Pd on charcoal]. The trisaccharide (1) thus obtained was purified by cellulose column chromatography with eluent A (EtOAc-PrⁱOH-H₂O; 3:3:2) (91%) [m.p. 143-148 °C (from MeOH-EtOAc); α_D^{20} + 36.5° (H₂O); homogeneous by silica gel t.l.c. (eluent A) and paper chromatography (eluent A: R_{lactose} 1.2, R_{fucose} 0.5; eluent, EtOAcpyridine-H₂O, 10:4:3: $R_{1actose}$ 1.0, R_{fucose} 0.28)]. N.m.r. data (240 MHz; D_2O ; Me_4Si in $CDCl_3$ as external reference) indicated the presence of a ca. 2:1 α , β -anomeric mixture: α -anomer: δ 5·40 (d, J 3·5 Hz, H-1 Gal), 5·14 (d, J 3·5 Hz, H-1 Fuc), 5.12 (d, J 3.7, H-1 GalNAc), 2.10 or 2.13 (s, NAc), and 1.26 or 1.30 (d, J 6 Hz, CMe); β -anomer: δ 4.73 (d, J 7.5 Hz, H-1 Gal), 5.30 (d, J 3.5 Hz, H-1 Fuc), 5.18 (d, J 3.7 Hz, H-1 GalNAc), 2.13 or 2.10 (NAc), and 1.30 or 1.26 (CMe). The presence of the N-acetyl galactosamine unit was confirmed by g.l.c. (SE 30 column at 180 °C) following acidic hydrolysis and trimethylsilylation.

The trisaccharide (1) inhibits hemagglutination of A_2 erythrocytes by human anti-A serum at the $80 \,\mu g/40 \,\mu l$ level. In this respect, and in its m.p., t.l.c., and paper

 \dagger When necessary, compounds were purified by silica gel column chromatography with the solvent mixture given as eluent. Optical rotations were determined for 1% solutions in CH₂Cl₂. All compounds exhibit satisfactory spectral and microanalytical properties.

[‡] The alcohol (5) may be recovered from the *cis* ether.

chromatography behaviour, it was identical with a natural sample isolated from galactosemic urine.11

The authors thank Dr. Cartron (Central National de Transfusion Sanguine) for the immunochemical study of synthetic (1) and Professor Montreuil (Université de Lille) for the gift of a sample of natural (1).

(Received, 13th March 1978; Com. 277.)

- ¹ W. M. Watkins, in 'Glycoproteins,' ed. A. Gottschalk, Elsevier, Amsterdam, 1972, p. 830.
 ² A. Guibal and C. Ropars, Rev. Fr. Transfusion Immuno-hematologie, 1976, 19, 127.
 ³ H. Paulsen, C. Kolar, and W. Stenzel, Angew. Chem. Internat. Edn., 1976, 15, 440.

- ⁹ H. Paulsen, C. Kolar, and W. Stenzel, Angew. Chem. Internat. Edn., 1976, 15, 440.
 ⁴ R. U. Lemieux, presented in part at the Spring Meeting of the Chemical Society Carbohydrate Group, Norwich, April 18—20, 1977.
 ⁶ P. A. Gent and R. Gigg, J.C.S. Perkin I, 1974, 1446.
 ⁶ S. David, J. Eustache, and A. Lubineau, J.C.S. Perkin I, 1974, 2274.
 ⁷ E. J. Corey and J. W. Suggs, J. Org. Chem., 1973, 38, 3224.
 ⁸ R. U. Lemieux, K. B. Hendriks, R. V. Stick, and K. James, J. Amer. Chem. Soc., 1975, 97, 4056.
 ⁹ S. David, A. Lubineau, and J. M. Vatèle, J.C.S. Perkin I, 1976, 1831.
 ¹⁰ S. David and A. Lubineau, Nouveau Journal de Chimie, 1977, 1, 375.
 ¹¹ G. Strecker T. Riazi-Ferzad B. Fournet S. Souguelet and I. Montreuil Biochimie, 1975, 58, 815.

- ¹¹ G. Strecker, T. Riazi-Ferzad, B. Fournet, S. Bouquelet, and J. Montreuil, Biochimie, 1975, 58, 815.